



RAPID'sakazakii – A Short Protocol Method for the Detection of Cronobacter spp. in Powdered Infant Formula Milk

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Introduction

Cronobacter spp. are Gram-negative, rod-shaped, motile and facultative-anaerobic bacteria originally listed as yellow-pigmented *Enterobacter cloacae*. The bacteria were subsequently called *Enterobacter sakazakii*. Taxonomic studies have determined that *E. sakazakii* comprises a high genetic heterogeneity and should be classified as a novel genus: *Cronobacter* ⁽²⁾. *Cronobacter* spp. has been implicated in severe infections such as meningitis, necrotizing enterocolitis, bacteremia and sepsis in neonates ⁽⁴⁾. In several of these cases, consumption of contaminated powdered infant formula milk PIFM was identified

as the source of *Cronobacter* ingestion ^(3,5). In response to this hazard, the International Dairy Federation (IDF) and the International Standard Organization (ISO) have standardized the reference method ISO/TS 22964 for the detection of *Cronobacter* spp. which includes of a double enrichment in buffered peptone water (BPW) and modified lauryl sulphate tryptose broth (mLST) followed by isolation on a selective chromogenic agar ⁽¹⁾. To meet this standard the RAPID'sakazakii agar has been developed for the detection of *Cronobacter* spp.

Its principle is based on the chromogenic detection of α-D-glucosidase common to all *Cronobacter* species. Hydrolysis of the chromogenic substrate leads to the formation of typical blue to turquoise colonies of *Cronobacter*. A shortened protocol consisting in direct plating on RAPID'sakazakii after a single enrichment in BPW has been proposed for the detection of *Cronobacter* spp. in powdered infant formula. The aim of this study was to assess the RAPID'sakazakii short protocol method compared to the double enrichment with the current ISO/TS 22964 standard (Figure 1).

Materials and Methods

All experiments were performed according to technical specifications of the EN ISO 16140 validation standard regarding inclusivity, exclusivity and the comparison study of methods. Inclusivity, the ability of the RAPID'sakazakii method to detect *Cronobacter* spp. with typical colonies was investigated with 24 *Cronobacter* strains (due to the recent change in nomenclature,

most strains are always designated *Enterobacter sakazakii*). Exclusivity, the ability of the method to distinguish non-target strains from *Cronobacter* spp. was tested with 34 strains of *Enterobacteriaceae*. The RAPID'sakazakii short protocol method was also evaluated in comparison with the reference method ISO/TS 22964 by

testing 62 powdered infant formula samples including 1 naturally contaminated sample and 30 artificially contaminated samples (6 samples by cross-contamination and 24 samples using a strain of *Cronobacter* stressed by heat treatment at 56°C for 15 minutes).

Results and Discussion

Figure 1: ISO/TS 22964 and RAPID'sakazakii protocols

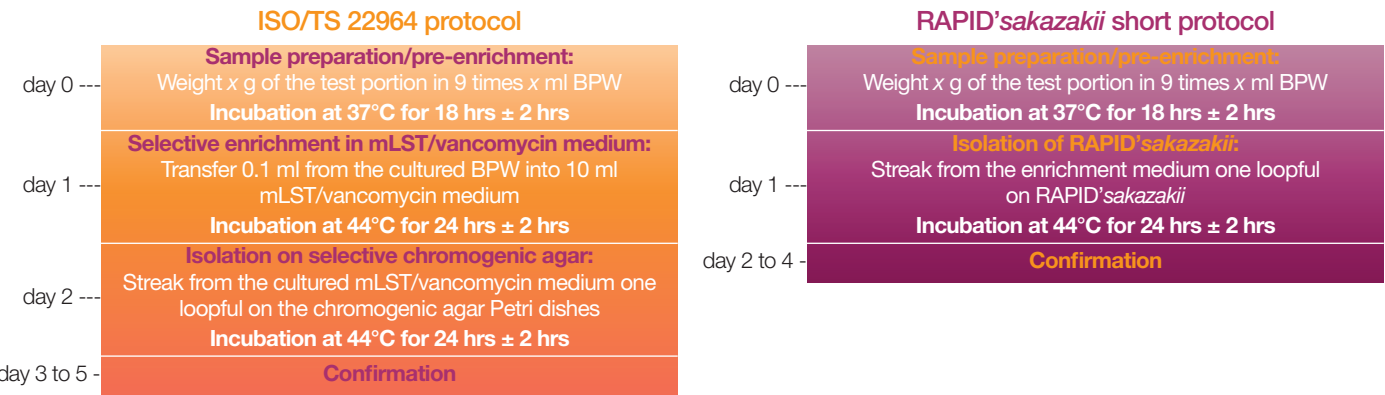


Table 1: Inclusivity study

Microorganism	ID	Growth	Colony color
<i>Enterobacter sakazakii</i>	IPL05/07/019	++	turquoise
<i>Enterobacter sakazakii</i>	IPL06/09/040	++	turquoise
<i>Enterobacter sakazakii</i>	IPL06/26/005	++	turquoise
<i>Enterobacter sakazakii</i>	IPL449076	++	turquoise
<i>Enterobacter sakazakii</i>	IPL480189	++	turquoise
<i>Enterobacter sakazakii</i>	AdriaX36	++	turquoise
<i>Enterobacter sakazakii</i>	AdriaX37	++	turquoise
<i>Enterobacter sakazakii</i>	AdriaX39	++	turquoise
<i>Enterobacter sakazakii</i>	AdriaX40	++	turquoise
<i>Enterobacter sakazakii</i>	AdriaX41	++	turquoise
<i>Enterobacter sakazakii</i>	RDC236	++	turquoise
<i>Enterobacter sakazakii</i>	RDC361	++	turquoise
<i>Enterobacter sakazakii</i>	RDC362	++	turquoise
<i>Enterobacter sakazakii</i>	RDC363	++	turquoise
<i>Enterobacter sakazakii</i>	RDC364	++	turquoise
<i>Enterobacter sakazakii</i>	RDC365	++	turquoise
<i>Enterobacter sakazakii</i>	RDC366	++	turquoise
<i>Enterobacter sakazakii</i>	RDC367	++	turquoise
<i>Enterobacter sakazakii</i>	LMG5740	++	turquoise
<i>Enterobacter sakazakii</i>	ATCC51329	++	turquoise
<i>Cronobacter sakazakii</i>	LMG23826T	++	turquoise
<i>Cronobacter dublinensis</i>	DSM 18705	++	turquoise
<i>Cronobacter malonaticus</i>	DSM18702	+	turquoise
<i>Cronobacter muytjensii</i>	CIP103581	+	turquoise

IPL: Strain from Institut Pasteur, France
Adria, Ad: Adria Collection, France
RDC: Bio-Rad R&D Collection, France

Table 2: Exclusivity study

Microorganism	ID	Growth	Colony color
<i>Enterobacter aerogenes</i>	ATCC13048	++	purple
<i>Enterobacter aerogenes</i>	RDC227	++	purple
<i>Enterobacter aerogenes</i>	Ad889	+/-	Faint purple
<i>Enterobacter agglomerans</i>	Adria11	+/-	faint purple
<i>Enterobacter amnigenus</i>	RDC222	-	/
<i>Enterobacter amnigenus</i>	Adria129	-	/
<i>Enterobacter amnigenus</i>	A00C068	-	/
<i>Enterobacter cloacae</i>	LMG2742	++	purple
<i>Enterobacter cloacae</i>	RDC22	++	purple
<i>Enterobacter cloacae</i>	Adria10	-	/
<i>Enterobacter cloacae</i>	RDC225	++	purple
<i>Enterobacter fergusonii</i>	Adria2876	++	faint purple
<i>Enterobacter gergoviae</i>	LMG5739	+/-	purple
<i>Enterobacter gergoviae</i>	CIP76.1T	-	/
<i>Enterobacter intermedius</i>	RDC234	-	/
<i>Enterobacter intermedius</i>	Adria60	-	/
<i>Enterobacter hormaechei</i>	Ad990	++	faint purple
<i>Enterobacter kobei</i>	RDC221	+	purple
<i>Enterobacter kobei</i>	Ad706	+/-	purple
<i>Enterobacter spp</i>	RDC205	++	purple
<i>Citrobacter freundii</i>	ATCC8090	+	purple
<i>Citrobacter koseri</i>	RDC186	+/-	purple
<i>Escherichia coli</i>	ATCC25922	++	grey blue to purple
<i>Escherichia coli</i>	DSMZ1576	++	grey blue to purple
<i>Escherichia hermannii</i>	RDC72	++	purple
<i>Escherichia vulneris</i>	RDC195	-	/
<i>Haefnia alvei</i>	CIP57.31	-	/
<i>Klebsiella oxytoca</i>	RDC30	++	opalescent blue
<i>Klebsiella pneumoniae</i>	ATCC13883	++	opalescent blue
<i>Leclercia adecarboxylata</i>	Ad707	-	/
<i>Pantoea agglomerans</i>	LMG1286T	-	/
<i>Salmonella Enteritidis</i>	ATCC13076	++	purple
<i>Salmonella Typhimurium</i>	ATCC14028	++	purple
<i>Serratia marcescens</i>	ATCC8100	-	/

Inclusivity and exclusivity

In pure culture testing, all *Cronobacter/E. sakazakii* strains produced typical turquoise colonies within 22 hours of incubation (Table 1). Most of the strains showed large colonies with a shimmering, highly visible turquoise color. None of the non-target strains gave typical colonies. Among 34 strains of *Enterobacteriaceae* tested, 2 strains belonging to the genus *Klebsiella* produced opalescent blue colonies easily distinguishable from *Cronobacter* colonies (Table 2).

Methods comparison study

Among 62 samples tested for *Cronobacter* spp. detection, 29 positive results were found with at least one method. Each method detected 30 positive samples and failed to detect 1 positive sample (Table 3).

Table 3: Paired results of the reference method ISO/TS 22964 and the RAPID'sakazakii method

	ISO/TS 22964 meth. +	ISO/TS 22964 meth. -
RAPID'sakazakii meth. +	29*	1**
RAPID'sakazakii meth. -	1†	31††

* Positive Agreement (PA) ** Positive Deviation (PD)
† Negative Deviation (ND) †† Negative Agreement (NA)

These results enabled the calculation of relative accuracy, relative specificity and relative sensitivity of the RAPID'sakazakii short protocol method compared to the reference method ISO/TS 22964 (Table 4).

Table 4: Relative accuracy, relative specificity and relative sensitivity of the RAPID'sakazakii method compared to ISO/TS 22964

	Rel. accuracy	Rel. specificity	Rel. sensitivity
Response to <i>Cronobacter</i> spp.	96.8%	96.9%	96.7%

The sensitivities of both methods, by taking into account all obtained positive results, were re-evaluated according to the EN ISO 16140 standard (Table 5).

Table 5: Sensitivities of the reference ISO/TS 22964 and the RAPID'sakazakii methods

	Ref. method	RAPID'sakazakii meth.
Response to <i>Cronobacter</i> spp.	96.7%	96.7%

Both methods provided exactly the same results for the detection of *Cronobacter* in powdered infant formulas.

Conclusion

The RAPID'sakazakii short protocol method was evaluated for the detection of *Cronobacter* spp. compared to the standardized reference method ISO/TS 22964. The inclusivity/exclusivity study demonstrated the superior specificity of the RAPID'sakazakii method. Regarding sensitivity, the performances of the RAPID'sakazakii method were similar to the reference method. Removing the 2nd enrichment step of the reference method, the

RAPID'sakazakii method reduces the time to result. Moreover, *Cronobacter* colonies are often larger and more colorful on RAPID'sakazakii agar compared to the reference chromogenic agar. In conclusion, the RAPID'sakazakii short protocol method has been demonstrated to be an efficient, convenient, cost and time-saving method for the detection of *Cronobacter* spp. in powdered infant formula milk.

References:

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